**LIPS Assay Protocol for Lassa Virus**

1. Dissolve 2 protease inhibitor tablets in 10ml of IP10 buffer (Label IP10+)

***Forcontrols (ELISA)***

1. Add 71.6µl of IP10+ buffer + 3.4µl of lysate(NP) per well
2. Transfer 25mls of each controls to each well

***For Samples***

1. Add 94.6µl of IP10+ buffer + 3.4µl of lysate(NP) per well
2. Transfer 2mls of each sample to each well
3. Incubate for 1hr at 250c at 180rpm
4. Multiply 5µl of A/G resin required for each well by the number of wells to be used. Measure and transfer the total amount into a 1.5ml eppendorf tube. Mark where the volume is.
5. Wash with 500µl of IP10 and centrifuge at 10,000 rpm for 3mins, remove supernatant and repeat 2 more times. Remove final supernatant and make up to the original volume.
6. Add 5µl/well of A/G ultrabeads to multiscreen plate

**(Note: Use the cut end of a p20 pippette tip)**

1. Transfer up to 130µl of the incubated sample + IP10+ into each well of the multiscreen plate containing the beads.
2. Incubate for 1hr at 250c at 180rpm
3. Use the vacuum pump to wash plate 8x with IP10 using 100µl/well
4. Use the vacuum pump to wash plate 2x with 1xPBS using 100µl/well
5. Blot the plate dry on paper towel
6. Add 50 µl/well of prepared luciferase substrate

**Note: To prepare luciferase substrate, for 96 wells, add 52 µl of substrate to 5200 µl of substrate buffer. Mix properly**

1. Incubate at room temperature for 10mins
2. Read plate on the Glomax.